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1       **Improvement of the performance and microstructural development of**  
2                               **alkali-activated slag blends**

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16  
17       **Abstract:** This study investigated the performance and microstructural  
18       development of alkali-activated slag (AAS) blends containing hydromagnesite  
19       seeds. AAS blends with and without seeds were assessed by isothermal  
20       calorimetry, porosity, setting time and compressive strength measurements. The  
21       formation of hydrate phases was further investigated via XRD, TGA-DSC, FTIR  
22       and SEM analyses. AAS blends incorporating 2% hydromagnesite seeds revealed  
23       the best performance in terms of hydration kinetics, strength and microstructural  
24       development. The positive influence of seeds was highlighted via increased  
25       hydration rates and degrees, shortened setting times, higher compressive  
26       strengths and denser microstructures. These improvements were attributed to the  
27       provision of additional nucleation sites in the presence of seeds that enabled the  
28       increased formation of hydrate phases (e.g. C-(A)-S-H with more crosslinks  
29       forming between adjacent silicate chains) within the pore structure.

31 **Keywords:** Alkali activated cement; kinetics; calcium-silicate-hydrate;  
32 performance; microstructure

## 1. Introduction

Alkali-activated cements (AACs) are being investigated as an alternative to ordinary Portland cement (OPC), due to the improvements they offer in terms of long-term chemical resistance and lower environmental impact associated during their production [1-3]. Recent studies looked into the use of AACs in foamed products [4] and 3D printed composites [5, 6]. Ground granulated blast-furnace slag (GGBS), produced by quenching molten iron slag from a blast furnace, has been widely used since the emergence of AACs [1]. Different activators such as sodium hydroxide and sodium silicate provide a high pH and thereby increase the solubility and dissolution of slag. This activation process facilitates the formation of hydrate phases such as C-(A)-S-H [7], resulting in a high mechanical performance and chemical resistance [1]. However, some drawbacks of AACs are still present, such as their susceptibility to drying shrinkage at room temperature, whereas recent studies [8] reported a reduction in total drying shrinkage with the inclusion of certain additives such as polyethylene glycol.

AACs can be considered under calcium- and silica-based and aluminosilicate-based systems. The rate and degree of hydration and the morphology of phases forming in AAS blends determine their mechanical properties due to the formation of different phases [2, 9]. Similar to the hydration of OPC, the hydration of slag under a high pH environment (e.g.  $> 12$ ) involves the stages of dissolution, induction, acceleration, deceleration and steady state diffusion [10]. However, during this process, the initially formed hydrate phases could create a layer and cover the surface of unreacted slag particles, which slows down the reaction and leads to a lower overall hydration degree.

One possible solution to this limitation in hydration involves the use of seeds in the initial mix design. These seeds can act as nuclei for the increased precipitation of hydrate phases. Because of its influence in accelerating the hydration kinetics of cementitious materials, nucleation seeding can be an effective approach in

enhancing the formation of hydrate phases at early ages [11]. A few studies investigated the effect of seeding in cementitious systems. These involved the use of synthesized C-S-H seeds as nucleation sites in OPC and pure tricalcium silicate ( $C_3S$ ) pastes [12], as well as AAS blends [13]. The obtained results revealed the benefits of seeding in the acceleration of hydration kinetics via the increased nucleation and growth of hydrate phases, which was previously identified as the rate limiting step during the hydration process. This led to an improved strength development at both early and later stages [12, 13].

Alternatively, recent studies [14] reported that while the synthesized C-S-H seeds played a role in the further precipitation of C-(A)-S-H, they did not necessarily facilitate the formation of the geopolymer gel. To obtain the C-S-H gel that is introduced into the mix for seeding purposes, the main method followed in the literature involves its synthesis through the reaction of sodium silicate and calcium nitrate [15] or calcium hydroxide and nano-silica [16] in a laboratory environment. This process is composed of several steps involving the washing, filtering and drying of the synthesized material, which is time consuming and is prone to carbonation when exposed to air [15]. Furthermore, differences in the Ca/Si ratio of C-S-H can influence the formation and properties of hydrate phases in the mixes it is introduced in [16], thereby limiting its use on a large scale in practice.

In addition to C-S-H, some nano particles were also involved as nucleation seeds due to their large surface areas [11, 17]. For instance, the use of nano-silica in OPC increased the hydration degree and rate, which was determined by the measurement of total surface area and calcium concentration [17]. Another study [18] revealed the catalytic effects of nano- $Al_2O_3$  particles in geopolymer mixes, which enabled an increased amount of gel phase formation within a short period of time due to their large surface areas. In addition to nano- $Al_2O_3$ , the nucleation and filling effects of nano- $CaCO_3$  and nano- $SiO_2$  were also reported in PC-based mixes that revealed a more compact and homogeneous structure [19]. Similarly, the use of nano clay [20] and nano metakaolin [21] also contributed to the strength

development of PC mixes. A similar effect was found in samples containing polycarboxylates, which can be an alternative admixture for improving the strength of concrete mixes [22]. Further studies [23-25] indicated that seeding was also effective in enhancing the hydration process in calcium-free binder systems. However, nano materials can pose a threat to human health and have a very high cost, presenting a challenge in their use in large scale applications [26, 27].

Other than nano materials, the use of hydromagnesite ( $4\text{MgCO}_3 \cdot \text{Mg}(\text{OH})_2 \cdot 4\text{H}_2\text{O}$ ) as an additive in cement-based mixes has also been investigated [23, 28]. Other studies [29] looked into the influence of  $\text{Mg}^{2+}$  on the kinetics of calcite precipitation and morphology. While it can be synthetically produced [30, 31], hydromagnesite is a naturally occurring mineral and is related to other magnesium containing minerals such as serpentine, dolomite and brucite [32]. Furthermore, its lower commercial cost than many of the nano materials such as nano-  $\text{SiO}_2$ ,  $\text{TiO}_2$  and  $\text{Al}_2\text{O}_3$  presents a benefit for the use of hydromagnesite. Unlike the hydration of pure  $\text{MgO}$  that leads to the formation of brucite, which does not result in a significant strength gain, a boost in the compressive strength of  $\text{MgO}$  mixes was seen in the presence of hydromagnesite [33]. Within these  $\text{MgO}$ -hydromagnesite blends, the formation of an amorphous phase was observed, which could contribute to their cohesive binding [34]. A similar outcome was observed in AAS mixes, where the combined use of  $\text{MgO}$  and hydromagnesite led to improved performance via the adjustment of the pH to accelerate the dissolution of GGBS [28].

Differing from the existing literature, this paper focuses on the incorporation of various amounts of hydromagnesite in AAS blends with the goal of identifying its effect on hydration kinetics and associated performance, which has not been investigated before. To reveal the role of hydromagnesite seeds in the improvement of reaction mechanisms, sample performance and microstructural development, the seeded samples were compared with the precursor (i.e. slag blend without any activator or seed) and unseeded AAS blends via the use of

different analysis techniques. The effect of nucleation seeding was evaluated by isothermal calorimetry, setting time, porosity and compressive strength measurements. X-ray diffraction (XRD), thermogravimetric analysis and differential scanning calorimetry (TGA-DSC), Fourier transform infrared spectroscopy (FTIR) with spectral subtraction and deconvolution, and scanning electron microscopy (SEM) were used in the analysis of the hydration process and the assessment of the amount and morphology of hydrate phases in both seeded and unseeded samples.

## **2. Materials and Methodology**

### **2.1 Materials and sample preparation**

The GGBS used in this study, whose chemical composition (i.e. obtained from the supplier, Engro Corporation Ltd. (Singapore)) is shown in Table 1, was grade P8000, referring to a super-reactive ultrafine slag powder (average particle size: < 5  $\mu\text{m}$  and fineness: 800-900  $\text{m}^2/\text{kg}$ , which is 2-3 times higher than common GGBS). In line with the findings of a previous study [13], the activator used was sodium metasilicate pentahydrate ( $\text{NaSiO}_3 \cdot 5\text{H}_2\text{O}$ , > 97%), obtained from VWR Pte. Ltd. (Singapore). The activator was included at 10% of the mass of the binder (slag). In terms of seeds, hydromagnesite ( $4\text{MgCO}_3 \cdot \text{Mg}(\text{OH})_2 \cdot 4\text{H}_2\text{O}$ , specific surface area (SSA): 43.5  $\text{m}^2/\text{g}$ ) was obtained from Fisher Scientific Ltd. (UK). Different amounts of hydromagnesite, corresponding to 1, 2 and 5% of the mass of the binder component, were used. The morphology of hydromagnesite presented in Fig. 1 demonstrated a rosette-like structure, which was in line with those reported in previous studies [35-38].

The prepared fresh mixture was cast into 50×50×50 mm cubic molds, consolidated by a vibrating table and finished by a trowel. All samples were demolded after 24

hours and cured under ambient temperature ( $30\pm1.5^{\circ}\text{C}$ ) and relative humidity ( $80\pm5\%$ ).

## **2.2 Methodology**

### **2.2.1 Isothermal calorimetry**

The heat flow corresponding to the hydration of AAS blends was investigated at  $30^{\circ}\text{C}$  by an I-Cal 2000 high precision calorimeter according to ASTM C1702-17 [39]. To prepare the paste samples, hydromagnesite seeds were first dispersed in the  $\text{NaSiO}_3\cdot 5\text{H}_2\text{O}$  solution by an ultrasonic homogenizer (Q500 sonicator), followed by the mixing of the slag into the solution, which was previously heated to  $30^{\circ}\text{C}$  (i.e. room temperature). Once the prepared pastes were ready, they were immediately placed into the calorimeter to record the heat released for up to 36 hours.

### **2.2.2 Setting time**

The setting time of AAS blends was evaluated by the measurement of the penetration depth of the Vicat needle into the prepared pastes in accordance with ASTM C191-08 [40]. The initial setting time was calculated to the nearest 1 minute according to Equation 1, where E represented the time (minutes) of the last penetration greater than 25 mm, H represented the time (minutes) of the first penetration less than 25 mm, C represented the penetration reading at time E and D represented the penetration reading at time H. The final setting time, calculated to the nearest 5 minutes, was determined by the time that elapsed between the initial contact of water and cement and the time that the needle was unable to sink visibly.



187 Initial setting time =  $((H - E)/(C - D)) \cdot (C - 25) + E$  (1)

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190 **2.2.3 Porosity**

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192 X-ray computed tomography ( $\mu$ -CT), performed with a SkyScan1183 scanner, a  
193 camera, and a flat panel sensor, was employed to analyze the porosity of the  
194 prepared blends, including the open and closed pores. Unlike other methods such  
195 as Mercury Intrusion Porosimetry (MIP), where the pore distribution is highly  
196 influenced by the intrusion pressure,  $\mu$ -CT is capable of accurately revealing the  
197 pore volume and size distribution without damaging the sample [41]. To facilitate  
198 this, the specimens obtained from samples crushed during strength testing were  
199 fixed on a micro-positioning stage and scanned using a source voltage and current  
200 of 80 kV and 100  $\mu$ A, respectively. The analysis of porosity involved the process  
201 of image acquisition, reconstruction, thresholding, pore labelling and quantification.  
202 During image acquisition, a representative 2D projection of each sample (i.e. at a  
203 diameter of 25 mm and spatial resolution of 7  $\mu$ m) was selected. The thresholding,  
204 pore labelling and quantification of porosity were conducted with the CT-Analyzer  
205 software.

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208 **2.2.4 Compressive strength**

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210 The compressive strength of AAS blends was tested at 3, 7 and 14 days. The test  
211 was conducted using a compression machine (Toni Technik Baustoffprüfssysteme  
212 GmbH), operated at a constant loading rate of 100 kN/min. The average and  
213 standard deviation of three samples were reported for each data point.

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216 **2.2.5 XRD, TGA-DSC, FTIR and SEM analyses**

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Hardened paste specimens extracted from the cubic samples crushed after strength testing were stored in acetone to stop hydration and dried under vacuum for 24 hours to prepare for microstructural analysis. Prior to microstructural analysis, the vacuum dried samples were ground down to pass through a 75  $\mu\text{m}$  sieve.

XRD was recorded on a Philips PW 1800 spectrometer using Cu  $K_{\alpha}$  radiation (40 kV, 30 mA) with a scanning rate of  $2^{\circ}$   $2\theta$ /step from  $5^{\circ}$  to  $80^{\circ}$  ( $2\theta$ ). TGA-DSC was conducted on a Perkin Elmer TGA 4000 equipment from 30 to  $900^{\circ}\text{C}$  with a heating rate of  $10^{\circ}\text{C}/\text{min}$  under nitrogen flow.

A Thermo Scientific Nicolet iS50 spectrometer with a built-in attenuated total reflection (ATR) module was used to obtain Fourier transform infrared spectroscopy (FTIR) spectra and identify the chemical components within the samples. Prior to analysis in the transmittance mode, the preparation of a pellet was conducted consisting of a mixture of 1 mg of powder from the samples with 300 mg of KBr. The mixture was pressed using 7 metric tons of hydraulic pressure in a die. In the ATR mode, powder samples were placed directly on the diamond ATR sampling station and pressed against the diamond crystal by a constant compressive force of 267 N. The resolution was set at  $4\text{ cm}^{-1}$  and a collection of 64 scans was obtained for each spectrum. Spectral subtraction and deconvolution of FTIR were also performed in line with the procedure explained in [42].

Before SEM analysis, samples were vacuum dried for 24 hours and coated with platinum under a 30 mA current, with a coating time of 30 seconds. The coated samples were then studied via a field emission scanning electron microscope (FESEM, 207 JOEL JSM-7600F). Backscattered electron (BSE) imaging was also used in SEM. To facilitate this, the vacuum dried samples were mounted in epoxy resin for 24 hours and polished to obtain smooth surfaces. Initial polishing was performed with a sandpaper before final polishing by diamond slurry with sizes of 9, 3 and  $0.3\text{ }\mu\text{m}$ . Dust and diamond particles generated during the polishing

process were removed by exposing the samples to an ultrasonic bath for 10 minutes, followed by vacuum drying for another 24 hours before the analysis.

#### **2.2.6 pH test**

The pH values of the activator and hydromagnesite seeds were measured via a calibrated Mettler Toledo pH meter, in line with ASTM C25 [43]. Accordingly, 5 grams of powder was blended with 100 grams of distilled water for 30 min prior to each test.

### **3. Results**

#### **3.1 Isothermal calorimetry**

The heat flow curves showing the reaction of AAS blends containing different amounts (i.e. 1-5% of the binder content by mass) of hydromagnesite during the first 36 hours are presented in Fig. 2. The hydration of GGBS by itself did not release any noticeable amount of heat, as indicated by the relatively flat heat curve. This was attributed to the low pH of this system, limiting the dissolution of the binder. Alternatively, when activated by the high pH provided by  $\text{Na}_2\text{SiO}_3 \cdot 5\text{H}_2\text{O}$ , all AAS blends indicated notable hydration peaks, albeit at different times and intensities. The common aspect of all AAS blends was the three stages of the hydration process. The first stage was the pre-induction stage, associated with the dissolution of slag [44]. The length and the timing of this stage depended on the type of the seed. The second stage was the induction period, which was significantly shortened due to the availability of a large number of nuclei provided by the presence of the seeds. The third stage of hydration was the nucleation, growth and precipitation of the hydrate phases. When compared to the unseeded blends, the advances in the hydration of blends containing seeds were associated

with the provision of additional nucleation sites supplied by the seeds within the pore space, away from the particle surfaces [12].

Increasing the amount of hydromagnesite seed from 1% to 2% led to an obvious reduction in the induction period and enabled the acceleration of the hydration reaction. A further increase in the seed content to 5% delayed the appearance of the main hydration peak when compared to blends containing 2% seed. This adverse effect could be associated with the agglomeration of the seed when its quantity was increased, which was in line with the observations reported in previous studies [24]. Accordingly, the high amount of seeds used in these blends could have led to the trapping of the alkali solution within the solid particles, thereby limiting the dissolution of the main binder phase. Therefore, hydromagnesite seed included at 2% of the overall binder content was chosen in the preparation of further mixes, whose details are shown in Table 2.

### **3.2 Setting time**

The initial and final setting time of all samples are presented in Fig. 3. The setting of all mixes generally took place during the acceleration period. In line with the isothermal calorimetry results, the sample without any activator or seed (GA0H0) demonstrated a very long setting time, which was associated with the initially low alkalinity of the pore solution. The incorporation of 10% activator in sample GA10H0 led to an obvious reduction in both the initial and the final setting times due to the increase in the pH of the solution, which enabled the dissolution of the binder and the precipitation of hydrate phases responsible for setting. The use of seeds along with the activator in sample GA10H2 had a much more significant effect on setting time, which was largely reduced. This notable reduction in the setting time was associated with the provision of nucleation sites via the addition of hydromagnesite. This led to an elimination of the induction period, thus reducing the setting time. Furthermore, the presence of hydromagnesite, which acted as

nuclei within the pore solution, favored the formation of a higher amount of hydrate phases, which led to the rapid hardening of sample GA10H2. These findings were in line with those of previous studies [45], where it was indicated that the incorporation of fine carbonate fillers led to a rapid increase in viscosity due to the improvement of the packing density of cement-based systems, resulting in shorter setting times in seeded samples.

### **3.3 Porosity**

The typical 2D images of cross-sections (i.e. diameter = 25 mm; spatial resolution = 7  $\mu\text{m}$ ) and corresponding porosities (i.e. shown in the lower left corner of each image) of all samples after 14 days of curing are presented in Fig. 4. Since  $\mu\text{-CT}$  analysis is known to enable the identification and quantification of both closed and open pores within the samples [46], the obtained results were a clear indication of the significant decline in porosity facilitated by the introduction of seeds in the sample GA10H2, where the volume taken up by solid phases was notably increased. This effect was attributed to the seeding effect of hydromagnesite particles, which enabled the abundant formation of hydrate phases within the pore solution. This could be further supported with the micro-filler effect of the seed itself, whose physical presence reduced the void space between large particles [47], albeit at a much smaller scale than the seeding effect. The micro-filler effect was not identified as the major cause of porosity reduction as the porosity of sample GA10H2 was 3.5 times smaller than that of sample GA10H0 (i.e. 16.9% vs. 59.7%), in the presence of only 2% hydromagnesite seeds by weight.

### **3.4 Compressive strength**

Fig. 5 presents the early strength development of all samples up to 14 days. All samples revealed a stable increase in strength with curing age due to the

continuation of hydration. Activated (GA10H0) and seeded (GA10H2) samples revealed higher 3-day strengths (17.1-19.1 MPa) than the control sample (GA0H0, 11.9 MPa). The notable strength of the control sample without the use of any activator or seeds could be associated with the relatively high reactivity and fineness of the GGBS used in this study. A similar trend was also observed at longer curing periods, during which the effect of seeding became more prominent. In line with the hydration kinetics and porosity results, the introduction of 10% of activator in sample GA10H0 led to ~30% improvement in strength at 14 days due to the increase in the dissolution of slag in the alkaline environment. While the 3-day strengths of samples GA10H0 and GA10H2 were comparable, an increase in the curing age resulted in an obvious improvement in the strength of the seeded sample. Accordingly, sample GA10H2 revealed a 14-day strength of 38.3 MPa, which was 71.7% higher than that of sample GA10H0 (22.3 MPa). This improvement in the presence of seeds was associated with the formation of a continuous network of hydrate phases around the solid particles within the pore solution. Without the addition of seeds, the precipitation of hydrate phases was restricted by the limited content of solid phases within the slag, whose surface area gradually reduced as hydrate phases started growing around them. Alternatively, the provision of additional nucleation sites enabled the further deposition of hydrate phases on the nuclei as well, which was reflected as improved performance in sample GA10H2 when compared to other samples. This could be further enhanced with the filler effect of the seed, which contributed to the densification of sample microstructure, and hence increased the strength in the long term.

### 3.5 XRD

Fig. 6 presents the XRD patterns of all samples after 14 days of curing. The broad hump located at around 25-35° 2 $\theta$  observed in sample GA0H0 was attributed to the presence of amorphous phases [48], whereas with the introduction of activator in samples GA10H0 and GA10H2 led to the appearance of C-(A)-S-H at 30-31° 2 $\theta$

(PDF #00-033-0306) as the major hydrate phase, which was in line with the findings of previous studies [49]. The XRD pattern of sample GA10H2 also demonstrated the presence of hydrotalcite ( $\text{Mg}_6\text{Al}_2(\text{OH})_{16}\text{CO}_3 \cdot 4\text{H}_2\text{O}$ ; PDF #01-070-2151), however at a lower intensity when compared with C-(A)-S-H. The  $\text{Mg}^{2+}$  source in hydrotalcite could be due to the dissolution of the slag. Alternatively, the presence of calcite in samples GA10H0 and GA10H2 ( $\text{CaCO}_3$ ; PDF #01-071-3699) could be attributed to the reaction between  $\text{Ca}^{2+}$  (i.e. from the dissolution of slag) and atmospheric  $\text{CO}_2$  via natural carbonation (Equation 2), albeit small due to the low concentration of  $\text{CO}_2$  in the atmosphere ( $\sim 0.04\%$ ). The absence of any obvious hydromagnesite peaks in the XRD patterns was due to the low contents of hydromagnesite (i.e. 2% of the binder content) present in the seeded samples, as well as the precipitation of hydrate phases around the seed particles. Overall, the XRD patterns confirmed that the formation of hydrate phases such as C-(A)-S-H was associated with the strength development of the seeded samples.



### 3.6 TGA-DSC

The TGA-DSC results of all samples after 14 days curing are shown in Fig. 7. The following three stages of thermal decomposition were observed:

(i) 30-220°C: Mass loss due to the dehydration of C-(A)-S-H, with an endothermic peak at  $\sim 80^\circ\text{C}$  [49].

(ii) 220-730°C: Mass loss due to the decomposition of hydrotalcite [49, 50].

(iii) 730-900°C: Mass loss due to the decarbonation of carbonates (e.g. calcite), with an endothermic peak at  $\sim 870^\circ\text{C}$  [24, 51].

The summary of mass loss values obtained at various stages of thermal decomposition after 14 days of curing is shown in Table 3. The bound water content calculated from the mass loss in C-(A)-S-H could be used as an assessment of the hydration degree in cement blends [50, 52]. In line with the XRD results, the introduction of seeds in sample GA10H2 revealed ~130% increase in the mass loss when compared with unseeded samples. Furthermore, the sharper endothermic peak observed at ~80°C in sample GA10H2 also hinted the presence of a higher amount of C-(A)-S-H than in other samples. Similarly, a slightly higher amount of mass loss corresponding to the decomposition of hydrotalcite was observed in sample GA10H2. The overall decomposition patterns revealed by all samples highlighted the higher contents of hydrate phases, especially C-(A)-S-H, in samples involving the simultaneous use of activator and seeds. These outcomes were in line with the XRD patterns, as well as the isothermal calorimetry, porosity and strength measurements presented earlier.

### 3.7 FTIR

The FTIR spectra of the seed (hydromagnesite) and all samples after 14 days of curing are shown in Fig. 8(a). The spectra of samples GA10H0 and GA10H2 revealed a peak at 800-1100  $\text{cm}^{-1}$ , which differed in shape and location (i.e. peak center) when compared with sample GA0H0. This observation was attributed to the stretching vibrations of Si-O [53, 54] due to the formation of C-(A)-S-H gel. Furthermore, a low-intensity band around 590-600  $\text{cm}^{-1}$  was demonstrated by samples GA10H0 and GA10H2, indicating the presence of C-A-S-H with the incorporation of Al into C-S-H [55].

In order to identify the FTIR peaks of the hydrate phases in samples GA10H0 and GA10H2, FTIR spectral subtraction [42] was conducted via subtracting the spectrum of the control sample (GA0H0) from the spectra of GA10H0 and GA10H2, respectively (Fig. 8(b)). As the band between 1080-1180  $\text{cm}^{-1}$  was mainly due to



the presence of unhydrated binder [42], the presence of hydrate phases could be conservatively defined through spectral subtraction. The spectra of hydrate phases in samples GA10H0 and GA10H2 (i.e. with baseline correction) were further investigated by spectral deconvolution [42] to reveal the individual bands hidden in the broad peaks. Fig. 9 presents these deconvolution results, which were used to indicate the Q values of Si-O. In terms of various spectra assignments, the peak around  $860\text{ cm}^{-1}$  was assigned to  $Q^1$  [56], whereas bands at  $900$  and  $950\text{-}980\text{ cm}^{-1}$  were unanimously assigned to stretching vibrations of  $Q^2$  sites [57, 58]. In line with the findings of previous studies [42, 59, 60], the peaks at  $898$ ,  $947$  ( $945$ ) and  $985\text{ cm}^{-1}$  were assigned to  $Q^2$ , as multiple  $Q^2$  sites could coexist in C-(A)-S-H. The deconvoluted spectra indicated that  $Q^2$  was the major band in both spectra. In addition to the  $Q^1$  and  $Q^2$  assignments, the peak at  $1030\text{ cm}^{-1}$  also indicated the presence of crosslinked Si-O, which was allocated to  $Q^3$  [54] in C-A-S-H gels. Along with the stretching vibrations of Si-O, the  $872\text{ cm}^{-1}$  band at a position similar to the bending band of  $\text{CO}_3^{2-}$  was observed, which usually occurred at around  $875\text{ cm}^{-1}$  [58]. The noticeably higher peak intensities associated with C-(A)-S-H gel in sample GA10H2, as revealed by Fig. 9, were an indication of a higher yield of C-A-S-H when compared to those of GA10H0. This observation was in agreement with the XRD and TGA findings, where an increased formation of hydrate phases was revealed within sample GA10H2 involving the simultaneous use of activator and seeds.

Table 4 shows the FTIR spectral deconvolution of the hydrate phases in samples GA10H0 and GA10H2. While the total percentage of  $Q^2$  in the C-(A)-S-H gel observed within both samples were similar, a greater percentage of  $Q^3$  sites in sample GA10H2 was observed. This difference highlighted a higher degree of polymerization of the hydrate phase, referring to the presence of a more-crosslinked C-(A)-S-H gel (i.e. revealing an improved polymerization of the aluminosilicate skeleton) in sample GA10H2 when compared to sample GA10H0. Moving on from previous studies [61], where it was shown that the formation of crosslinked C-(A)-S-H was accountable for high strength gain in AAS blends, this

finding could also contribute to the understanding of the improved reaction mechanisms and associated performance of sample GA10H2.

### 3.8 SEM

Fig. 10 shows the microstructural images of selected samples after 14 days of curing. When compared with the unseeded sample (GA10H0), the introduction of hydromagnesite seeds in sample GA10H2 led to a denser microstructure, resulting in a considerable strength improvement in sample GA10H2, which was in line with the porosity results. The microstructural image of hydromagnesite, shown in Fig. 10(c), confirmed the growth of the hydrate phases (i.e. possibly C-(A)-S-H, as indicated by XRD and FTIR results) around the rosette-like hydromagnesite crystals, which grew and covered the surfaces of other particles. Previous studies [62] reported that the adsorption force between  $\text{Ca}^{2+}$  ions and calcite ( $\text{CaCO}_3$ ) was a moderately strong acid-base (donor-acceptor) interaction, identified as an ionic-covalent bond. This interaction was mainly related with the active surface sites resulting from the presence of  $\text{CO}_3^{2-}$  in calcite, although its solubility was relatively low [63]. Therefore, it can also be inferred that the interaction between  $\text{Ca}^{2+}$  ions and hydromagnesite could also be based on an ionic-covalent bond due to the presence of  $\text{CO}_3^{2-}$  in hydromagnesite ( $4\text{MgCO}_3 \cdot \text{Mg}(\text{OH})_2 \cdot 4\text{H}_2\text{O}$ ). Accordingly,  $\text{Ca}^{2+}$  ions were first chemically adsorbed via a strong force, which led to a reduction in their mobility, resulting in the formation of “stable nuclei”. This was followed by the precipitation of the C-(A)-S-H gel on the surface of these nuclei, during which  $\text{Ca}^{2+}$  ions could establish a connection between the formed C-(A)-S-H gel and the hydromagnesite particles by ionic-covalent bonds.

### 3.9 pH

In addition to its role as a nucleation seed, the dissolution of hydromagnesite may affect the pH of the solution and hence influence the hydration process. To eliminate any such concern, a comparison of the pH values of the activator solution with and without the addition of 2% hydromagnesite seeds was shown in Fig. 11. These measurements revealed stable pH values across the two solutions, indicating that hydromagnesite particles mainly acted as nucleation sites. This was also confirmed by the low pH of the pure hydromagnesite solution, showing that the majority of hydromagnesite particles remained undissolved when incorporated in the activator solution.

#### **4. Discussion**

Heterogeneous nucleation can occur through the seeding of the nuclei of new phases into the existing phases, due to a lower free energy barrier [64, 65]. Accordingly, two types of heterogeneous nucleation and associated growth of hydrate phases were expected to take place simultaneously within the seeded AAS samples prepared in this study: (i) Hydration that started on the surfaces of the original slag particles, and (ii) additional hydration that started on the surfaces of hydromagnesite seeds distributed amongst the large pores in AAS blends. In line with this expectation, Fig. 12 shows a schematic representation of the hydration process in samples GA10H0 and GA10H2. After several days of curing, unseeded samples (GA10H0) experienced a restriction of further nucleation and growth via the initial formation of a thick dense layer of hydrate phases, leaving a large quantity of capillary pores within the microstructure. This constraint led to a large porosity and inferior strength in comparison to seeded samples (GA10H2), in which the hydrate phases (e.g. C-(A)-S-H gel) nucleated both on the original particle surfaces as well as the seeds. When compared to the unseeded samples, the addition of seeds into AAS blends provided an increased amount of nuclei for the precipitation of additional hydrate phases, thereby greatly reducing the induction time for hydration. This advancement in the hydration process increased

the reaction kinetics, which was beneficial for strength development. The nucleation of hydrate phases between the pores not only accelerated the hydration rate and increased the overall hydration degree, but also led to denser microstructures. These findings were in line with the seeding mechanisms previously observed in OPC, C<sub>3</sub>S and alkali-activated slag systems [12, 13].

## **5. Conclusion**

This study demonstrated the effect of seeding in the acceleration of the hydration kinetics of AAS blends by investigating the role of different amounts of hydromagnesite as nucleation seeds. Within the seed content range (i.e. 1-5% of the binder content by mass) used in this study, the introduction of 2% hydromagnesite seeds proved to be effective in accelerating the hydration kinetics and increasing the overall hydration degree of the prepared blends. This was enabled via the continuation of hydration on the surface of the original binder as well as the seeds. The improvement observed in the hydration process led to fast setting within AAS blends, which translated into a dense microstructure via the widespread precipitation of hydrate phases (e.g. C-(A)-S-H). When compared to unseeded samples, the increase in the formation of crosslinked C-(A)-S-H was responsible for the high strengths achieved by the seeded samples.

Overall, the findings of this study highlighted the increased potential of AAS blends to be used in cement-based applications, considering their improved mechanical performance in the presence of a small amount of seeds. Its straightforward incorporation into the initial mix design, ease of application, lack of any safety concerns and lower cost when compared with alternatives (e.g. synthesized C-S-H and nano materials), have led to the identification of hydromagnesite as an ideal choice for enhancing the performance of AAS blends.

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559

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List of Tables

**Table 1** The chemical composition (wt.%) of GGBS.

	MgO	CaO	SiO <sub>2</sub>	Fe <sub>2</sub> O <sub>3</sub>	Al <sub>2</sub> O <sub>3</sub>	R <sub>2</sub> O <sub>3</sub>	K <sub>2</sub> O	TiO <sub>2</sub>
GGBS	9.3	37.8	32.3	0.5	14.6	-	-	-

**Table 2** The mix designs used in this study.

Sample	Activator content (%)	Hydromagnesite seed content (%)
GA0H0	0	0
GA10H0	10	0
GA10H2	10	2

**Table 3** Summary of mass loss obtained at various stages by TGA-DSC after 14 days of curing.

Sample	30-220°C	220-730°C	730-900°C	Total
	Dehydration of C-(A)-S-H	Decomposition of hydrotalcite	Decarbonation of carbonates	
GA0H0	4.2	3.7	1.3	9.2
GA10H0	5.4	5.1	1.0	11.5
GA10H2	12.4	7.0	0.7	20.1

**Table 4** FTIR spectral deconvolution of the hydrate phase in samples GA10H0 and GA10H2, provided as wavenumber (cm<sup>-1</sup>) and peak area percentage.

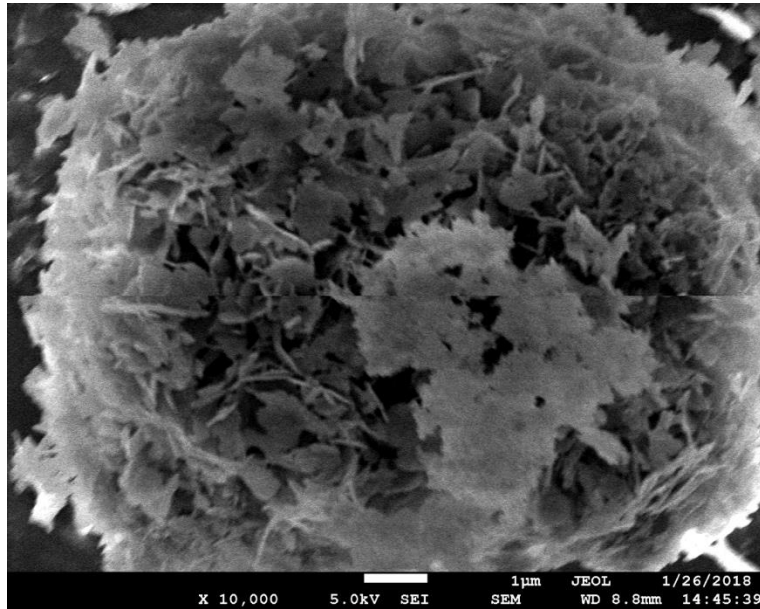
Band	1	2	3	4	5	6
Hydrate phase in sample GA10H0	840 (7.4%) <sup>a</sup>	-	898 (28.0%)	947 (35.4%)	985 (15.5%)	1030 (13.7%)
Hydrate phase in sample GA10H2	840 (4.3%)	872 (1.4%)	898 (25.1%)	945 (33.5%)	985 (18.8%)	1030 (17.0%)
Assignment <sup>b</sup>	Q <sup>1</sup>	CO <sub>3</sub> <sup>2-</sup>	Q <sup>2</sup>	Q <sup>2</sup>	Q <sup>2</sup>	Q <sup>3</sup>

<sup>a</sup> value in parentheses is the ratio of the component peak area to the broad area.

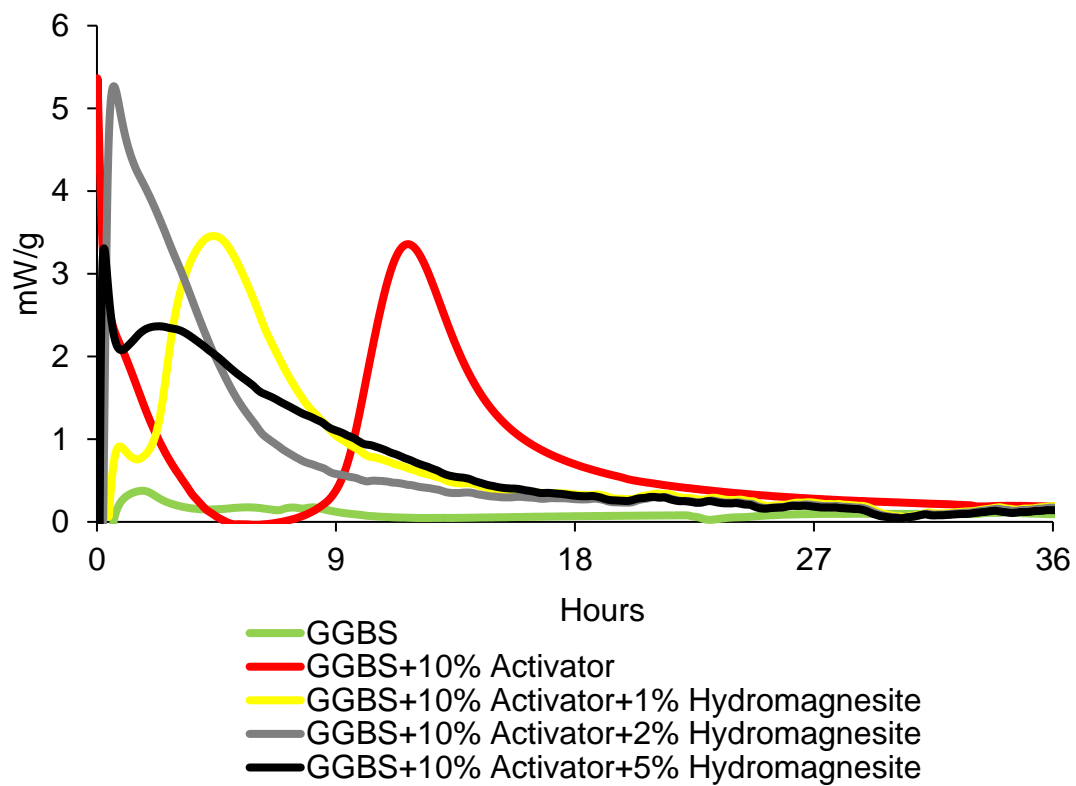
<sup>b</sup> Q denotes SiO<sub>4</sub> tetrahedron, whereas the superscripts indicate the number of other tetrahedron units attached to it.



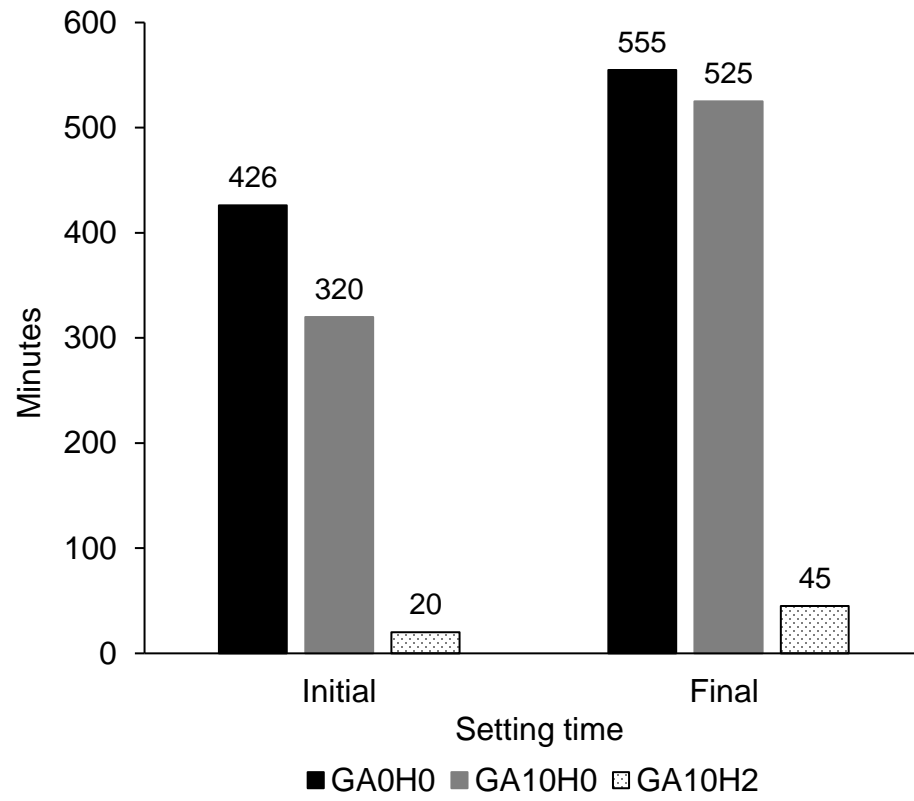
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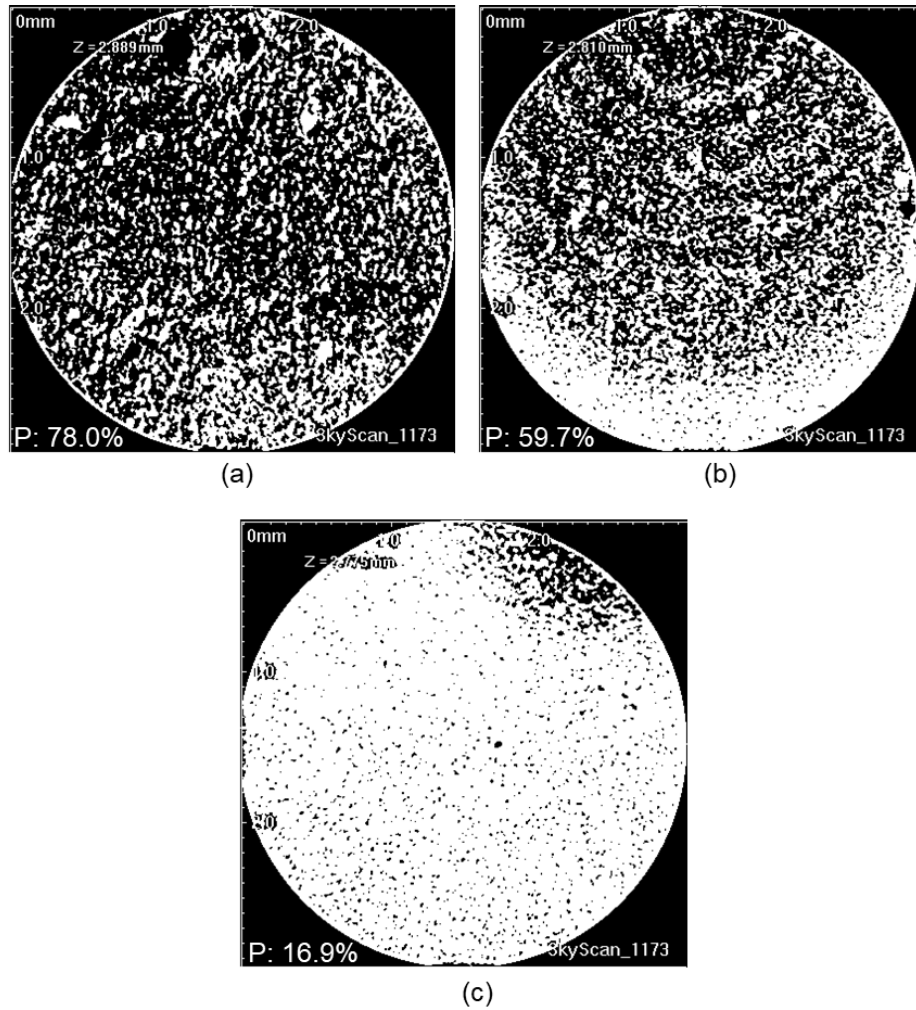
**Fig. 1** SEM image of the hydromagnesite seed used in this study



**Fig. 2** Heat flow curves of AAS blends containing different amounts of hydromagnesite seed

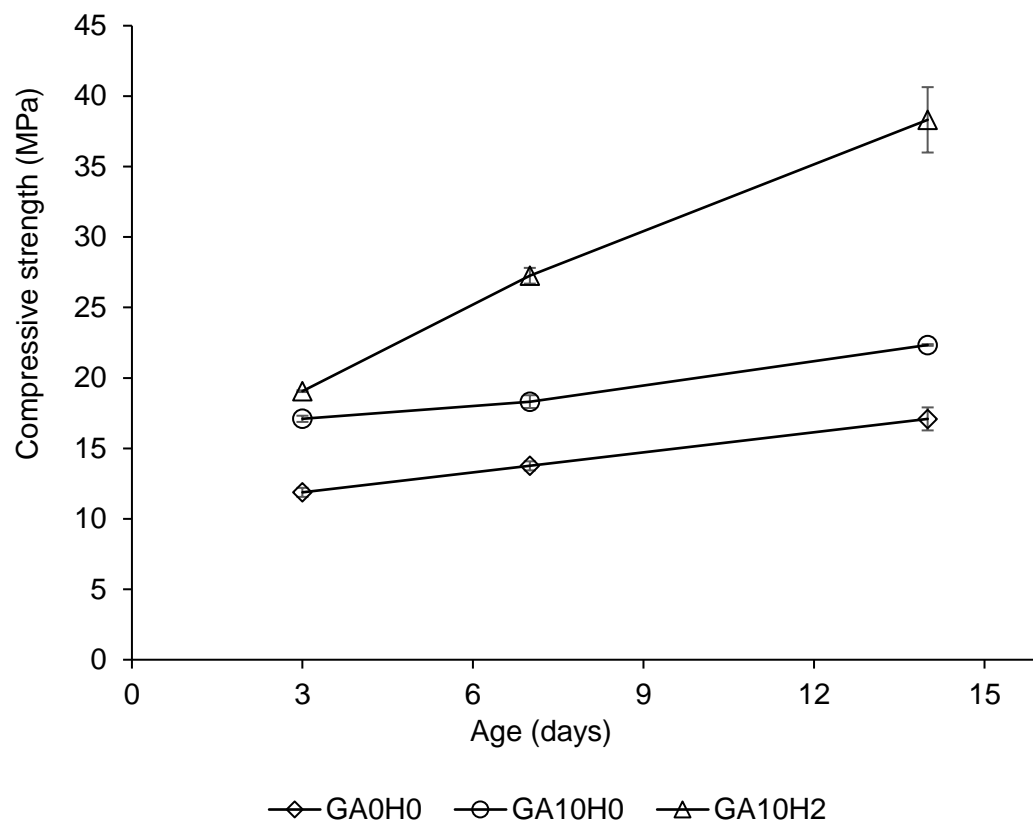


**Fig. 3** The setting time of samples with varying amounts of activator and seeds

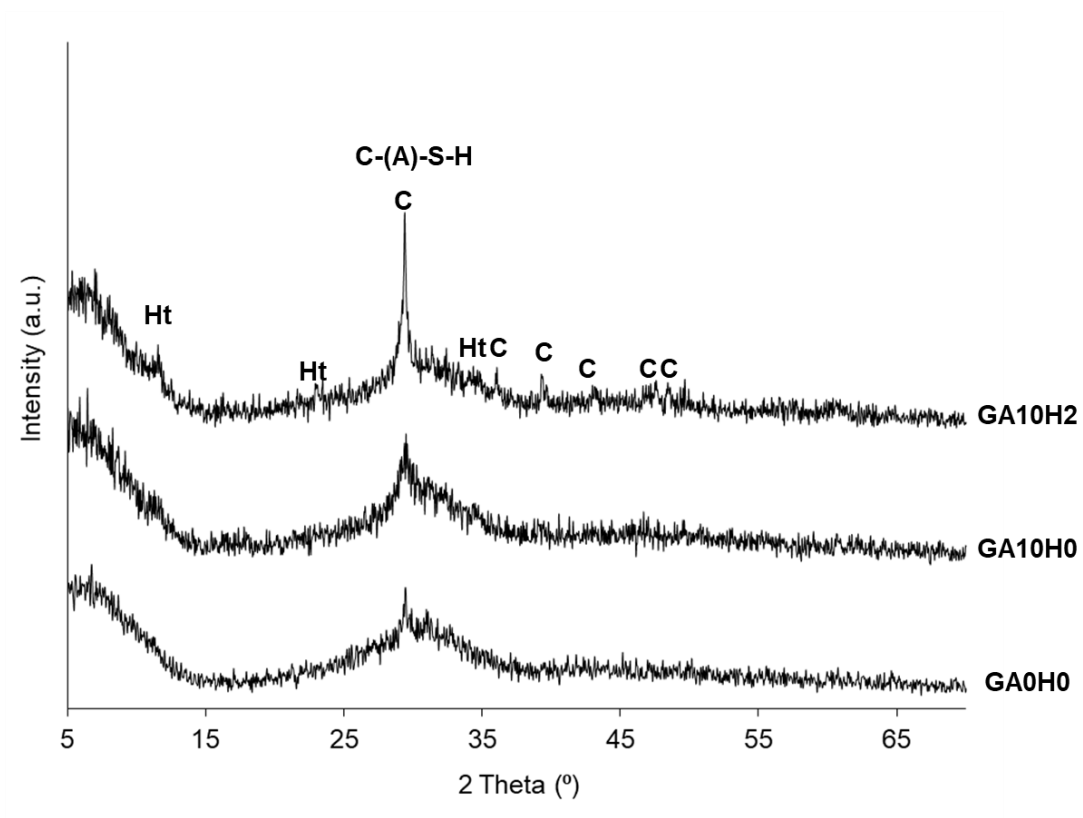


**Fig. 4** Typical 2D images of cross-sections (i.e. diameter = 25 mm, spatial resolution = 7  $\mu$ m) and corresponding porosities (i.e. shown in the lower left corner of each image) of samples (a) GA0H0, (b) GA10H0 and (c) GA10H2 after 14 days of curing

(white area: solid phases; black area: pores)

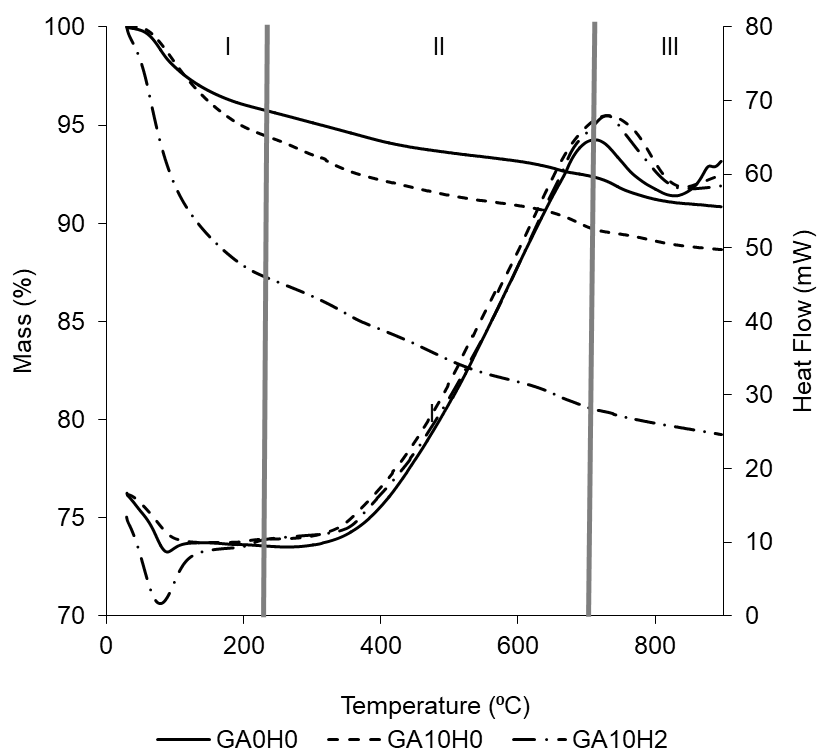


**Fig. 5** Compressive strength of samples with varying amounts of activator and seeds during 14 days of curing

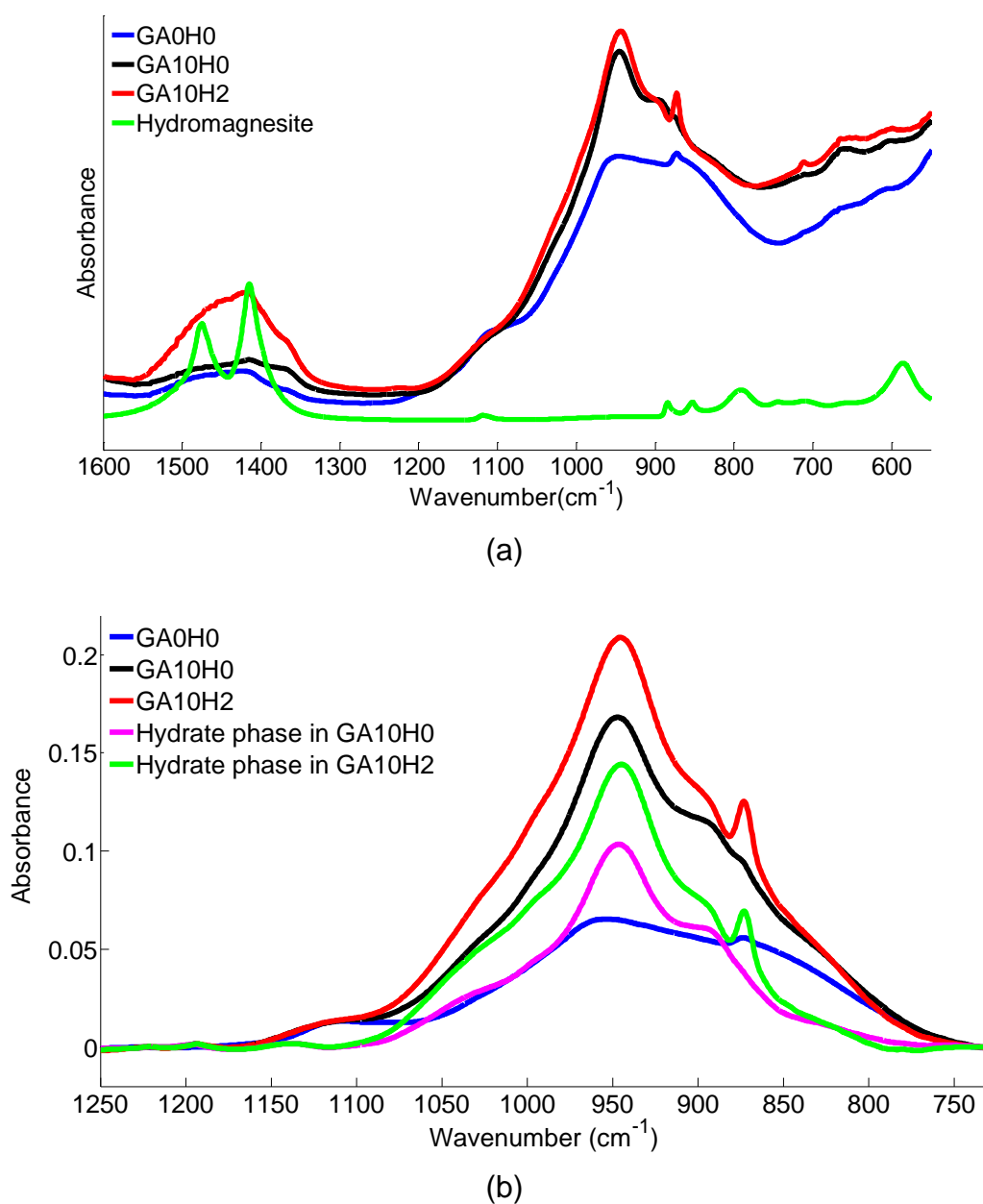


**Fig. 6** XRD pattern of samples with varying amounts of activator and seeds at 14 days

(Ht: Hydrotalcite; C: Calcite; C-(A)-S-H: Calcium-(aluminate)-silicate-hydrate)

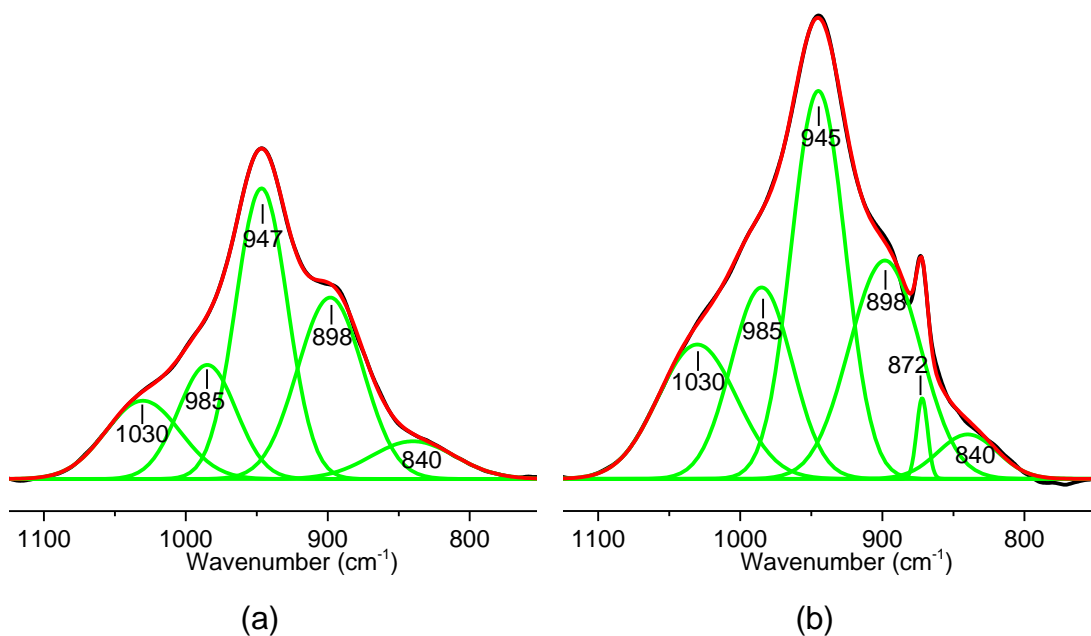


**Fig. 7** TGA-DSC results of samples with varying amounts of activator and seeds at 14 days



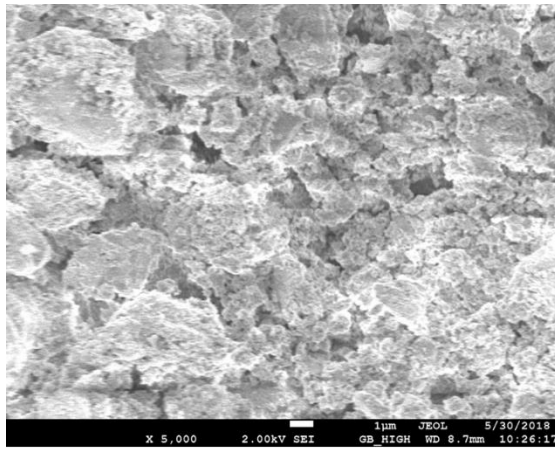
**Fig. 8** FTIR spectra of (a) hydromagnesite seed and samples with varying amounts of activator and seeds and (b) subtraction of the spectra of sample GA0H0 from samples GA10H0 and GA10H2 (i.e. after baseline correction) at 14 days



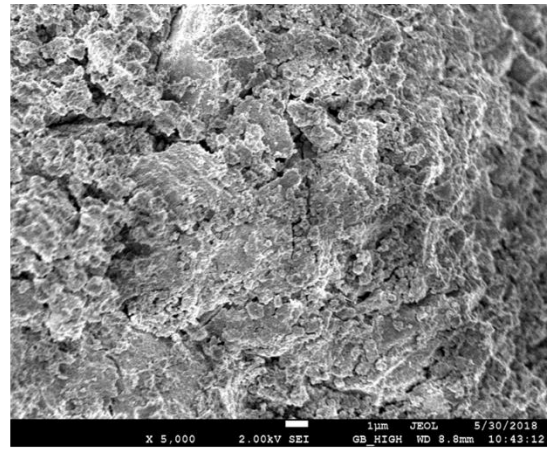


**Fig. 9** Deconvoluted FTIR spectra of hydration phases in samples (a) GA10H0 and (b) GA10H2

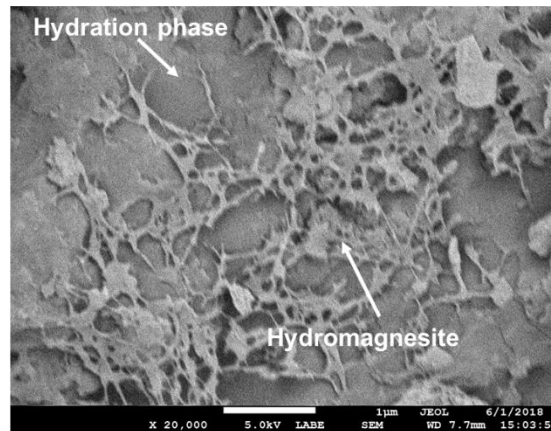
(black curve represents the original spectrum; green curves represent the bands associated with individual components; and red curve represents the sum of all the individual components)



(a)

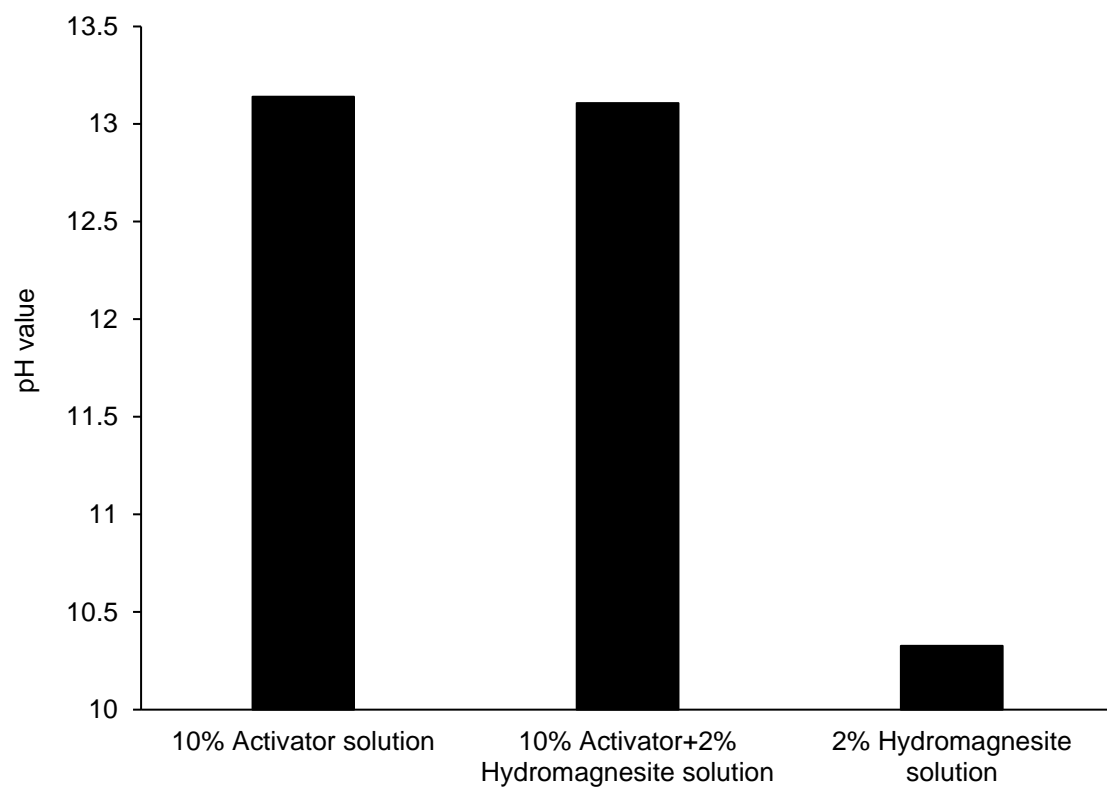


(b)

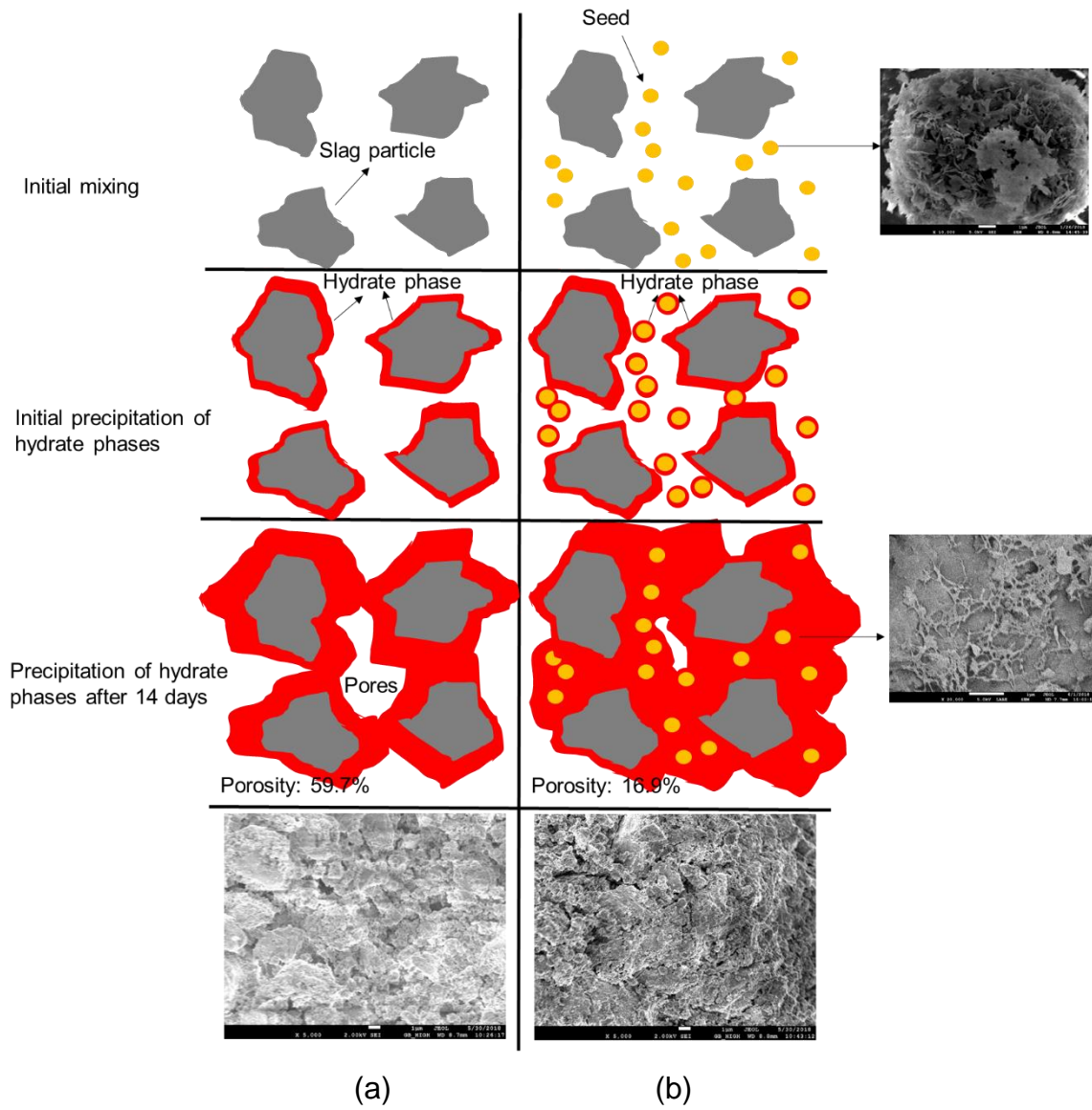


(c)

**Fig. 10** Microstructural images of (a) sample GA10H0, (b) sample GA10H2 and (c) the surface of a hydromagnesite particle at 14 days



**Fig. 11** pH values of different solutions used in this study



**Fig. 12** Schematic representation of the hydration process in samples (a) GA10H0 and (b) GA10H2